

ORIGINAL ARTICLE

Open Access

Effect of nano silver and silver nitrate on seed yield of (*Ocimum basilicum* L.)

Fatemeh Nejatzadeh-Barandozi^{1*}, Fariborz Darvishzadeh² and Ali Aminkhani³

Abstract

Background: The aim of this study was to evaluate the effect of nano silver and silver nitrate on yield of seed in basil plant. The study was carried out in a randomized block design with three replications.

Results: Four levels of either silver nitrate (0, 100, 200 and 300 ppm) or nano silver (0, 20, 40, and 60 ppm) were sprayed on basil plant at seed growth stage. The results showed that there was no significant difference between 100 ppm of silver nitrate and 60 ppm concentration of nano silver on the shoot silver concentration. However, increasing the concentration of silver nitrate from 100 to 300 ppm caused a decrease in seed yield. In contrast, a raise in the concentration of nano silver from 20 to 60 ppm has led to an improvement in the seed yield. Additionally, the lowest amount of seed yield was found with control plants.

Conclusions: Finally, with increasing level of silver nitrate, the polyphenol compound content was raised but the enhancing level of nano silver resulting in the reduction of these components. In conclusion, nano silver can be used instead of other compounds of silver.

Keywords: Basil; Nano silver; Silver nitrate; Seed yield; Polyphenol compounds

Background

Basil (*Ocimum basilicum* L.) is aromatic herbs that are used extensively to add a distinctive aroma and flavor to food. The leaves can be used fresh or dried as a spice. Essential oils extracted from fresh leaves and flowers can be used as aroma additives in food, pharmaceuticals, and cosmetics [1-3]. Traditionally, basil has been used as a medicinal plant in the treatment of headaches, coughs, diarrhea, constipation, warts, worms, and kidney mal-function [1]. Phytohormones and environmental stresses are the effective factors in controlling abscission process [4]. It is demonstrated that the ethylene has an important role in initiation of the abscission layer in different plants [5]. Ethylene activates the biosynthesis genes of hydrolytic enzymes, e.g., cellulose and polygalacturonase, which induces separation of plant organs from the main plant [6,7]. In addition, the abscission process could be regulated by the other phytohormones such as auxin (IAA) and abscisic acid (ABA). The later induces the abscission process through stimulation of the ethylene

biosynthesis while auxin is effective in delaying the abscission by reducing the sensitivity of cell to ethylene [4,6,8]. Abscission may be delayed by using some chemical components such as amino isobutyric acid and cobalt ions, (aminoxy) acetic acid, silver thiosulfate (STS), and silver nitrate (AgNO_3) [8,9]. Several studies demonstrated that spraying of silver ions decreases the flowers and flower bud abscission in orchid plant [10]. Additionally, it has been reported that silver ions decreased 100% flower abscission of *Alstroemeria* plant as compared to untreated flower within two first days [11]. Moreover, ethylene is involved on senescence of flowers in *Bougainvillea* plant while this process may be postponed by spraying silver thiosulfate [12].

Nano silver solution consisting silver ions in the size range of 10 to 100 nm, and it has more stability in comparison to other solutions. Nano silver particles, also, have more surface area in contact to outer space due to their small size. Thus, the amount of adhesion to the cell surface is increasing which lead to their higher efficacy [13]. Additionally, nano silver may affect the metabolism, respiration, and reproduction of microorganism [14]. For example, the effect of nano silver on extend maintenance period of leaves (from 2 to 21 days) in asparagus plant is

* Correspondence: fnejatzadeh@yahoo.com

¹Department of Horticulture, Faculty of Agriculture, Khoy Branch, Islamic Azad University, P.O. Box 58168-44799, Khoy, Iran

Full list of author information is available at the end of the article

Table 1 Analysis of variance mean square testing traits

SOV	Df	LN	PH	DWP	DWI	LL	LW	SY	WS	PP	TN	SC
R	2	65.95 ns	34.01 ns	5446.50**	1401.62**	1.15 ns	1.85 ns	9.66**	0.060**	0.66*	0.13 ns	282.63*
T	6	315.84**	30.23*	3775.01**	794.02**	0.44 ns	0.23 ns	30.40**	0.044**	0.35*	0.26**	5888.46**
E	12	30.39	10.35	773.46	116.30	0.98	0.56	0.92	0.002	0.103	0.043	65.016
CV		10.20	8.64	9.40	5.70	10.14	14.22	4.63	3.40	11.78	11.7	9.06

* and ** respectively, significant levels of 5% and 1%. LN, leaf number; PH, plant height; DWP, dry weight of plant; DWI, dry weight of inflorescence; LL, leaf length; LW, leaf width; SY, seed yield; WS, weight of 100 seeds; PP, polyphenol; TN, tannin; SC, silver concentration.

reported. Also, during this period, the amount of ascorbate, chlorophyll, and fiber were more in treated leaves [15]. The effect of silver nitrate in delaying the abscission has been studied [15]; however, the influence of nano silver on seed abscission has not been reported. Therefore, this study was carried out to assess the possibility of using nano silver and silver nitrate in delaying the time of seed abscission in basil plant.

Methods

This study was carried out in a completely randomized block design with three replicates in the field research of Islamic Azad University of Khoy (Iran). The data of this study was analyzed by SAS software, and the comparison was done according to least significant difference (LSD) test method. Four levels of either silver nitrate (0, 100, 200, and 300 ppm) or nano silver (0, 20, 40, and 60 ppm) were sprayed with 0.1% Tween 20 (Tween 20, Sigma-Aldrich, St. Louis, MO, USA) on basil plant at seed growth stage (that is 45 days after cultivation) and were repeated after 2 weeks. The nano silver solution with an average particle diameter of 25 nm was obtained from Pars Nano Nasb Company (Pars Nano Nasb Company, Tehran, Iran). Different parameters (that is, leaf number, plant height, dry weight of plant, length and width of leaf, dry weight of inflorescence, seed yield, and weight of 100 seeds) were determined; greenness of leaves was measured by a chlorophyll meter. The

biochemical properties included polyphenol and tannin were determined according to [16]. Concentration of silver in the plant shoot was measured by 'inductively coupled plasma' according to the method described by [16]. The principal operating parameters of the instrument were as follows: argon gas flow: auxiliary, 1 L/min, nebulizer (crossflow), 0.8 L/min; sample uptake: 60 s. Measurements were carried out in the axial mode at 328.068 nm.

Results

Using silver (either as nano silver or silver nitrate) had a significant effect on silver concentration in the plant shoot, number of leaves, height of the plant, plant dry weight, inflorescence dry weight, seed yield, weight of 100 seeds, and polyphenol and tannin content in shoot (Table 1). In contrast, the length, width, and greenness of leaves have not affected by silver treatment (Table 1). Silver concentration in shoot was increased by all treatments (either nano silver or silver nitrate) However, the higher concentration of silver in the shoot was obtained by silver nitrate treatment in comparison to others (Figure 1). There was no significant difference between 100 ppm of silver nitrate and 60 ppm concentration of nano silver on the shoot silver concentration (Figure 1).

The highest number of leaves was obtained by spraying 100 ppm of silver nitrate, while the lowest was seen in control plants (Table 2). The number of leaves and plant

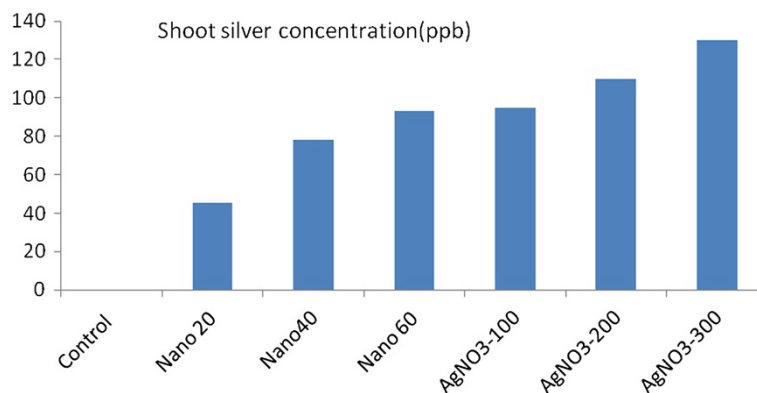


Figure 1 Effect of nano silver and silver nitrate treatments on concentration of silver. In plant tissue ($\mu\text{g/g}$ dry tissue of plant).

Table 2 Comparison mean effect of different concentrations of nano silver and silver nitrate on test traits

Treatment	LN	PH (cm)	DWP (g/m ²)	DWI (g/m ²)	LL (cm)	LW (cm)	SY (g/m ²)	WS (g)	PP	TN
Control	41.530e	30.977c	246.94d	161.90d	8.300a	5.040a	14.62e	2.01e	3.160a	2.220a
NS 20 ppm	46.24de	34.68abc	268.38 dc	174.68 cd	8.324a	5.367a	18.10d	1.860b	2.941ab	1.985ab
NS 40 ppm	50.06cde	38.392ab	301.50abc	188.10bc	8.501a	5.351a	20.41c	1.761bc	2.781abc	1.779bc
NS 60 ppm	53.364 cd	41.445a	347.06a	203.1ab	8.910a	5.701a	22.68b	1.70 cd	2.420ab	1.605 dc
NS 100 ppm	68.130a	39.810ab	325.79ab	210.45a	9.321a	5.725a	24.90a	1.615d	2.208c	1.265d
NS 200 ppm	67.431ab	7.025abc	303.72abc	199.8ab	9.090a	5.110a	23.14b	1.714 cd	2.595abc	1.672bc
NS 300 ppm	57.465bc	33.500bc	278.65bcd	189.30bc	9.920a	5.028a	19.71 cd	1.785bc	3.048a	1.960abc
LSD	9.965	5.750	49.660	19.268	1.765	1.350	1.725	0.105	0.570	0.376

Mean values, followed by the same letters in each column are not significantly different (Duncan's multiple range test at 5%). NS, nano silver; SN, silver nitrate; LN, leaf number; PH, plant height; DWP, dry weight of plant; DWI, dry weight of inflorescence; LL, leaf length; LW, leaf width; SY, seed yield; WS, weight of 100 seeds; PP, polyphenol, TN, tannin.

height was improved by increasing the concentration of nano silver while they were reduced by rising in silver nitrate concentration. The longest height of the plant was obtained with the treatment of nano silver 60 ppm, while the lowest amount of this feature was seen with control (Table 2). However, by going up, the concentration of silver nitrate from 100 to 300 ppm caused a decrease in the dry weight of plant and dry weight of inflorescence. In contrast, a raise in the concentration of nano silver from 20 to 60 ppm has led to an improvement in those parameters (Table 2). Raising the concentration of silver nitrate from 100 to 300 ppm caused a decline in the seed yield, but an increase in concentration of nano silver from 20 to

60 ppm leads to arise in the seed yield. The lowest amount of seed yield was found with control samples, and the highest was obtained by silver nitrate 100 ppm treatment. However, seed yield was found to be more than the control sample with different concentrations of nano silver and silver nitrate (Table 2). The highest weight of 100 seeds was observed in the control sample, while this weight was decreased when either nano silver or silver nitrate was applied (Table 2).

The highest content of polyphenol and tannin was observed in control, and the lowest content was observed with silver nitrate 100 ppm (Table 2). The polyphenol and tannin content was raised with the increasing level

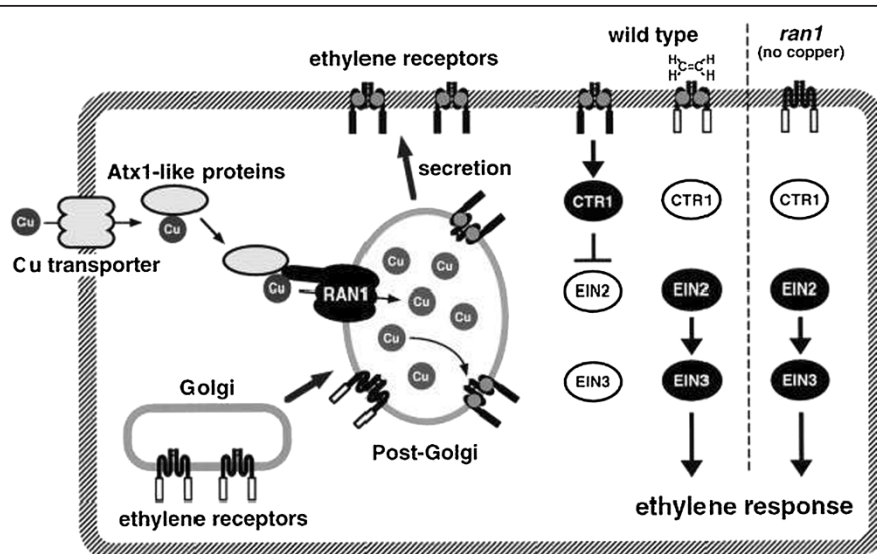


Figure 2 A model for the function of RAN1 in the ethylene signaling pathway. RAN1 is presumed to be localized in the membrane of a post-Golgi compartment. Copper ions received from CCH, a putative copper chaperon, is transported by RAN1 into a post-Golgi compartment, delivering the metal to membrane-targeted ethylene receptor apoproteins that become able to coordinate ethylene after the incorporation of copper ions. In the absence of the hormone, the receptors are active and negatively regulate downstream signaling components, preventing hormone-response phenotypes. Ethylene is expected to inactivate the receptors upon binding, presumably by causing a reduction in histidine kinase/phosphatase activity. This, in turn, results in derepression of downstream signaling components (EIN2, EIN3) and activation of hormone-response phenotypes. The metal-deficient ethylene receptors are nonfunctional, resulting in a constitutively activated signaling pathway.

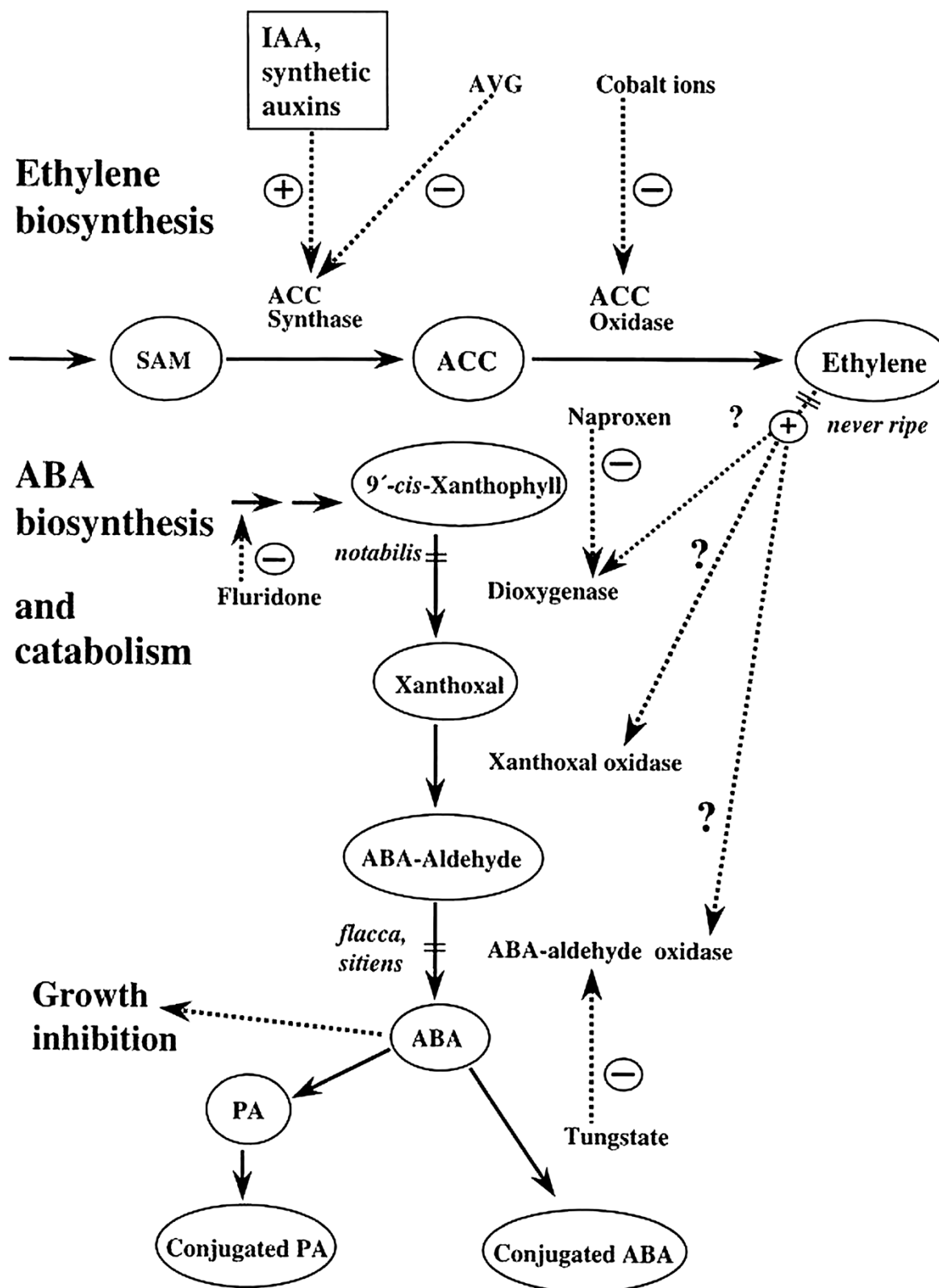


Figure 3 Inhibit biosynthesis of ethylene. Target sites of inhibitors (dotted arrows) are indicated. SAM, S-adenosyl-methionine.

of silver nitrate but was decreased with the increase in nano silver. The greenness, length, and width of the leaves were not affected by nano silver and silver nitrate treatments (Table 1).

Discussion

Application of silver ions can displace copper ions from the receptor proteins (Figure 2) consequently, block ethylene perception, since copper ions have a critical role in ethylene binding upon receptors [17-19]. This effect of silver ion on ethylene was reported by several researchers [11,20,21]. Therefore, some results in this study can be referred to the effect of silver in preventing the ethylene action (Figure 3). Decrease of plant's height due to ethylene is proven. For example, mutants of tobacco and *Arabidopsis*, which synthesize a high concentration of ethylene, have lesser height as compared to their wild species [22]. Furthermore, Nichols and Kofranek [23] reported that silver ion causes the increase in stem height of tulips and rose plants. Under the *in vitro* conditions, silver nitrate inhibited biosynthesis of ethylene and caused regeneration of multiple shoots from hypocotyl sections of cotton [24]. Another researcher reported that silver nitrate is effective in increasing wet weight of tobacco [25]. Ethylene caused an increase in enzyme activity of chlorophyllase and destruction of internal membrane of chloroplast, while that 100 ppm of silver nitrate caused decreasing the production of ethylene and destruction of chlorophyll in calamondin fruit [26]. Spraying silver nitrate causes improvement of seed yield in wheat [15]. In the present experiment, foliar application of either nano silver or silver nitrate causes increase in seed yield as compared to control (Table 2). In basil plant, rise in seed yield can be due to increase in the number of inflorescence in unit area, number of seed in the inflorescence, weight of seeds, and decrease in seed abscission. Considering that the seed yield has a significant positive correlation with dry weight of inflorescence ($r = 0.808$, $p < 0.0001$), but it has a significant negative correlation with weight of 100 seeds ($r = -0.835$, $p < 0.0001$). Consequently, it can be concluded that the increase of seed yield in this study is due to the increase of the inflorescence number in unit area and decrease in seed abscission.

Seed abscission is one of the main factors in reducing seed yield in basil plant. It is proven that one of the reasons for plant organ abscission is imbalance between phytohormones. Ethylene is playing an important role in this process. Furthermore, it is proven that silver ions inhibit the ethylene action by preventing its connection to its receptors in plant cells [8, 6, 4]. Thus, the increase in the seed yield was a result of reducing the seed abscission due to the inhibitory effect of silver on ethylene action. These results are confirmed by the results obtained from other studies [10,11,15,20]. There are many reports

about the use of silver nitrate in decreasing the abscission, but this study is the first research which is about nano silver effect on decreasing the abscission of reproductive organs of plants. Increasing silver concentration in aerial organs of sprayed plants with nano silver caused decreasing polyphenol and tannin content. While there was an increase in polyphenol and tannin content by spraying silver nitrate due to rising of the silver concentration in aerial organs of plant. There are many reports available about ethylene effect on increasing the phenol content [27,28]. On the other hand, high concentration of heavy metals (that is silver) in plant tissue causes polymerization of phenol by peroxidase enzyme which chelates the heavy metals [28,29]. Therefore, by increasing the level of silver nitrate more than 100 ppm, phenol and tannin content enhances due to toxicity effect of silver on plant cells.

Conclusions

There was no significant difference between 100 ppm of silver nitrate and 60 ppm concentration of nano silver on the shoot silver concentration. Therefore, permeability of nano silver is far greater than that of silver nitrate. The reason of this matter is the small size of nano particle, which causes more adhesion of nano particles to plant tissues. By considering the lesser use of silver in nano silver, this treatment can be used instead of other combinations of silver. However, the nano silver effect compared with other silver combinations on reducing the ethylene effect needs more researches.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Department of Horticulture, Faculty of Agriculture, Khoj Branch, Islamic Azad University, P.O. Box 58168-44799, Khoj, Iran. ²Department of Horticulture, Faculty of Agriculture, Garmsar Branch, Islamic Azad University, P.O. Box 58168-44799 Garmsar, Iran. ³Department of Chemistry, Khoj Branch, Islamic Azad University, P.O. Box 58168-44799, Khoj, Iran.

Received: 18 August 2014 Accepted: 12 September 2014

Published online: 02 October 2014

References

1. Simon JE, Morales MR, Phippen WB, Vieira RF, Hao Z (1999) A source of aroma compounds and a popular culinary and ornamental herb. In: Janick J (ed) Perspectives on new crops and new uses. ASHS Press, Alexandria, VA, pp 499-505
2. Javanmardi J, Khalighi A, Kashi A, Bais HP, Vivanco JM (2002) Chemical characterization of basil (*Ocimum basilicum* L.) found in local accessions and used in traditional medicines in Iran. *J Agri Food Chem* 50:5878-5882
3. Senatore F (1996) Influence of harvesting time on yield and composition of the essential oil of a thyme (*Thymus pulegioides* L.) growing wild in Campania (Southern Italy). *J Agri Food Chem* 44:1327-1332
4. Taylor J, Whitelaw C (2001) 'Singal in abscission' 2001, Tencley review no. 127, USA
5. Macnish AJ, Irvig D, Joyce DC, Vithanage V, Wearing AH (2004) Anatomy of ethylene-induced floral-organ abscission *Chamaelucium uncinatum* (Myrtaceae). University of Florida, Gainesvil, USA
6. Mishra A, Khare S, Trivedi PK, Nath P (2008) Effect of ethylene, 1-MCP, ABA and IAA on break strength, cellulose and polygalacturonase activities during cotton leaf abscission. *Afr J Bot* 282:6-12

7. Wu Y, Deng Y, Li Y (2008) Change in enzyme activities in abscission zone and berry drop of Kyoho grapes under high O₂ or CO₂ atmospheric storage. *LWT* 41:175–179
8. Kushad MM, Poovaiah BW (1984) Deferral of senescence and abscission by chemical inhibition of ethylene synthesis and action in bean explants. *Plant Physiol* 76:293–296
9. Moore TC (2006) *Biochemistry and physiology of plant hormones*, 2nd edn. Ferdowsi university press, Iran, pp 262–266
10. Uthaichay N, Ketso S, Van doorn WG (2007) 1-MCP pretreatment prevents bud and flower abscission in *Dendrobium* orchids. *Postharvest Boil Technol* 43:374–380
11. Wagstaff C, Chanasut U, Harren FJM, Laarhoven LJ, Thomas B, Rogers HJ, Stead AD (2005) Ethylene and flower longevity in *Alstroemeria*: relationship between tepal senescence, abscission and ethylene biosynthesis. *J Exp Bot* 56:1007–1016
12. Chang YS, Chen HC (2001) Variability between silver thiosulfate and 1-naphthaleneacetic acid applications in prolonging bract longevity of potted bougainvillea. *Scie Hort* 87:217–224
13. Shah V, Belozerova I (2008) Influence of metal nanoparticles on the soil microbial community and germination of lettuce seeds. *Water Air Soil Pollut* 4:9797–9799
14. Lok CN, Ho CM, Chen R, He QY, Yu WY, Sun H, Tam PKH, Chiu JF, Che CM (2007) Silver nanoparticles: partial oxidation and antibacterial activities. *Biol Inorg Chem* 12:527–534
15. Labraba X, Araus JL (1991) Effect of foliar applications of silver nitrate and ear removal on carbon dioxide assimilation in wheat flag leaves during grain filling. *Field Crops Res* 28:149–162
16. Makkar HSP (2000) Quantification of tannins in tree foliage. A laboratory manual for the FAO/IAEA co-ordinated research project on use of nuclear and related techniques to develop simple tannin assays for predicting and improving the safety and efficiency of feeding ruminants on tanniniferous tree foliage, FAO/IAEA Working Document, IAEA, Vienna, Austria
17. Khan NA (2006) *Ethylene action in plants*, 1st edn. Springer-Verlag Berlin Heidelberg, German
18. Means AR, Malley BWO, Armen LRH, Tashjian JR (2005) *Vitamins and hormones*. V. 72. Elsevier Inc, USA, pp 395–430
19. Hedden EP, Thomas SG (2006) *Annual plant reviews*. Blackwell Publishing, Oxford, UK, pp 125–139
20. Eo J, Lee BY (2009) Effects of ethylene, abscisic acid and auxin on fruit abscission in water dropwort (*Oenanthe stolonifera* DC.). *Scie Hort* 123:224–227
21. An J, Zhang M, Wang SH, Tang J (2008) Physical, chemical and microbiological change in stored green asparagus spears as affected by coating of silver nanoparticle-pvp. *LWT* 41:1100–1107
22. Pessaraki M (2001) *Handbook of plant and crop physiology*, 2nd edn. Marcel Dekker, Inc, USA, pp 35–108
23. Nichols R, Kofranek AM (1982) Reversal of ethylene inhibition of tulip stem elongation by silver thiosulfate. *Scie Hort* 17:71–79
24. Ouma JP, Young MM, Reichert NA (2004) Optimization of in vitro regeneration of multiple shoots from hypocotyl sections of cotton (*Gossypium hirsutum* L.). *Afr J Biotechnol* 3:169–173
25. Tso TC, Sorokin TP, Engelhaupt ME (1973) Effects of some rare elements on nicotine content of the tobacco. *Plant Physiol* 51:805–806
26. Purvis AC (1980) Sequence of chloroplast degreening in calamondin fruit as influenced by ethylene and AgNO₃. *Plant Physiol* 66:624–627
27. Park HJ, Kim SH, Kim HJ, Choi HS (2006) A new composition of nanosized silica-silver for control of various plant diseases. *Plant Pathol* 22:295–302
28. Heredia B, Cisneros-zevallos L (2009) The effects of exogenous ethylene and methyl jasmonate on the accumulation of phenolic antioxidants in selected whole and wounded fresh produce. *Food Chem* 115:1500–1508
29. Elzaawely AA, Xuan TD, Tawata S (2007) Changes in essential oil, kava pyrones and total phenolics of *Alpinia zerumbet* (Pers.) B.L. Burtt. and R.M. Sm. leaves exposed to copper sulphate. *Environ Exper Bot* 59:347–353

doi:10.1186/s13588-014-0011-0

Cite this article as: Nejatzadeh-Barandozi et al.: Effect of nano silver and silver nitrate on seed yield of (*Ocimum basilicum* L.). *Organic and Medicinal Chemistry Letters* 2014 **4**:11.

Submit your manuscript to a SpringerOpen[®] journal and benefit from:

- Convenient online submission
- Rigorous peer review
- Immediate publication on acceptance
- Open access: articles freely available online
- High visibility within the field
- Retaining the copyright to your article

Submit your next manuscript at ► springeropen.com